

Contents lists available at ScienceDirect

Vaccine

journal homepage: www.elsevier.com/locate/vaccine



Review

A model international partnership for community-based research on vaccine-preventable diseases: The Kamphaeng Phet-AFRIMS Virology Research Unit (KAVRU)



Robert V. Gibbons ^{a,1}, Ananda Nisalak ^{a,1}, In-Kyu Yoon ^{a,1}, Darunee Tannitisupawong ^{a,1}, Kamchai Rungsimunpaiboon ^b, David W. Vaughn ^{c,1}, Timothy P. Endy ^{d,1}, Bruce L. Innis ^{e,1}, Donald S. Burke ^{f,1}, Mammen P. Mammen Jr. ^{g,1}, Robert McNair Scott ^{h,1}, Stephen J. Thomas ^{i,1}, Charles H. Hoke Jr. ^{j,*,1}

- ^a Armed Forces Research Institute of Medical Science, Department of Virology, Bangkok, Thailand
- ^b Kamphaeng Phet Provincial Hospital, Kamphaeng Phet, Thailand
- ^c GlaxoSmithKline Vaccines, Rixensart, Belgium
- ^d Department of Infectious Diseases, State University of New York, Syracuse, NY, United States
- ^e GlaxoSmithKline Vaccines, King of Prussia, PA, United States
- f Graduate School of Public Health, University of Pittsburgh, Pittsburgh, PA, United States
- g Vical, Incorporated, San Diego, CA, United States
- h Bethesda, MD, United States
- ¹ Walter Reed Army Institute of Research, Silver Spring, MD, United States
- ^j Columbia, MD, United States

$A\ R\ T\ I\ C\ L\ E\quad I\ N\ F\ O$

Article history: Received 21 May 2013 Received in revised form 12 July 2013 Accepted 30 July 2013 Available online 8 August 2013

Keywords:
Vaccine efficacy trials
Site selection
Japanese encephalitis vaccine
Hepatitis A vaccine
Dengue vaccine
Influenza
Collaboration
Team building
Public Health

ABSTRACT

This paper describes an international collaboration to carry out studies that contributed to the understanding of pathogenesis, diagnosis, treatment, and prevention of several diseases of public health importance for Thailand and the United States. In Kamphaeng Phet Province, Thailand, febrile syndromes, including encephalitis, hepatitis, hemorrhagic fever, and influenza-like illnesses, occurred commonly and were clinically diagnosed, but the etiology was rarely confirmed. Since 1982, the Kamphaeng Phet Provincial Hospital, the Thai Ministry of Public Health, and the US Army Component of the Armed Forces Research Institute of Medical Sciences, along with vaccine manufacturers and universities, have collaborated on studies that evaluated and capitalized on improved diagnostic capabilities for infections caused by Japanese encephalitis, hepatitis A, dengue, and influenza viruses. The collaboration clarified clinical and epidemiological features of these infections and, in large clinical trials, demonstrated that vaccines against Japanese encephalitis and hepatitis A viruses were over 90% efficacious, supporting licensure of both vaccines. With the introduction of Japanese encephalitis vaccines in Thailand's Expanded Program on Immunization, reported encephalitis rates dropped substantially. Similarly, in the US, particularly in the military populations, rates of hepatitis A disease have dropped with the use of hepatitis A vaccine. Studies of the pathogenesis of dengue infections have increased understanding of the role of cellular immunity in responding to these infections, and epidemiological studies have prepared the province for studies of dengue vaccines. Approximately 80 publications resulted from this collaboration. Studies conducted in Kamphaeng Phet provided experience that contributed to clinical trials of hepatitis E and HIV vaccines, conducted elsewhere. To provide a base for continuing studies, The Kamphaeng Phet-AFRIMS Virology Research Unit (KAVRU) was established. This paper reviews the origins of the collaboration and the scientific observations made between 1982 and 2012.

© 2013 Elsevier Ltd. All rights reserved.

^{*} Corresponding author at: 6405 Cardinal Lane, Columbia, MD, United States. Tel.: +1 301 452 8897.

E-mail addresses: RobertVGibbons@gmail.com (R.V. Gibbons), AnandaN@afrims.org (A. Nisalak), Inkyu.yoon@afrims.org (I.-K. Yoon), DaruneeT@afrims.org (D. Tannitisu-pawong), kam.director1@gmail.com (K. Rungsimunpaiboon), david.w.vaughn@gsk.com (D.W. Vaughn), EndyT@upstate.edu (T.P. Endy), Bruce.2.Innis@GSK.com (B.L. Innis), DonBurke@pitt.edu (D.S. Burke), mammen.mammen@vical.com (M.P. Mammen Jr.), scottrmcn@cs.net (R.M. Scott), Stephen.J.Thomas3.mil@mail.mil (S.J. Thomas), CHHokeIr@gmail.com (C.H. Hoke Ir.).

¹ Author addresses: R.G., A.N., I.Y., D.T., D.V., T.E., B.I., D.B., M.M., S.T., and C.H. were assigned to the Armed Forces Research Institute of Medical Sciences (AFRIMS) when the work described in this paper was accomplished. RMS was assigned to the Walter Reed Army Unit Nepal (WARUN).

Contents

1.	Introduction					
2.	Histo	ry of the	collaboration	4489		
	2.1.	KPP Ho	spital	4489		
	2.2.	Thai Mi	nistry of Public Health	4489		
	2.3.	South E	ast Asia Treaty Organization (SEATO) Medical Research Laboratory (SMRL), Bangkok, Thailand (Fig. 1) [1]	4489		
	2.4.	The Arr	ned Forces Research Institute of Medical Sciences (AFRIMS)	4489		
	2.5.	Kamph	aeng Phet-AFRIMS Virology Research Unit (KAVRU)	4490		
3.	Scien	tific resul	ts of the AFRIMS-KPP collaboration (Table 1)	4490		
	3.1.	Japanes	e encephalitis (Fig. 2).	4490		
		3.1.1.	JE IgM antibody capture ELISA	4490		
		3.1.2.	JE vaccine in KPP	4490		
		3.1.3.	Entomological studies	4491		
	3.2.	Hepatit	is A studies (Fig. 3)	4491		
		3.2.1.	Invention of hepatitis A vaccine	4491		
		3.2.2.	Hepatitis A vaccine trial in KPP.	4492		
		3.2.3.	Post trial impacts	4492		
	3.3.	Dengue		4492		
		3.3.1.	Dengue diagnosis	4493		
		3.3.2.	Dengue pathogenesis (Fig. 4).	4493		
		3.3.3.	Dengue vaccine trials	4493		
		3.3.4.	Dengue vectors	4493		
	3.4.	Respira	tory and other disease studies	4494		
4.	Discu	ıssion		4494		
	4.1.	4.1. Site selection				
	4.2.					
		4.2.1.	Shared experience	4494		
		4.2.2.	Applicability of scientific results in KPP to development of vaccines against a range of tropical infectious diseases	4494		
		4.2.3.	Medical benefits	4494		
		4.2.4.	Community benefits	4494		
		4.2.5.	Challenges and positive/negative aspects of the conduct of the trials and lessons learned from conducting these			
			studies (Table 2)	4495		
	4.3.	Future :	studies	4495		
5.						
	Funding					
	Discl	Disclaimer				
	Acknowledgements (Table 3)					
	Refer	rences		4497		

1. Introduction

Vaccine efficacy trials are best conducted in areas where the incidence and epidemiology of the disease of interest is well-documented and recognized as important by local public health, medical and political leaders, as well as the community at-large. Of additional importance are the willingness of subjects to participate, their access to medical care and laboratory diagnostic facilities, and the clinical trials infrastructure supporting the various aspects of trial design and execution.

Less tangible are mutual respect, trust and long-term commitment between investigators, and these factors are particularly important for studies conducted jointly by host-country and visiting foreign investigators in which professional relationships are as important as formal institutional agreements.

We review a successful collaboration between investigators at Kamphaeng Phet Province (KPP) Hospital, Thailand, the Ministry of Public Health (MOPH) of Thailand, the US Army Medical Component-Armed Forces Research Institute of Medical Sciences (AFRIMS) in Bangkok, and other organizations. The collaboration took place in Kamphaeng Phet Province (KPP) in northern Thailand. The province was defined as a political entity before 1000 AD, and ruins from this period have been declared a UNESCO World Heritage Site. The province is generally hilly, though flat portions support the cultivation of rice, bananas, sugar cane, and tapioca, irrigated by the Ping River. The province covers 8600 km² subdivided into 11 districts containing just over 800 villages; the 2011 population was 726,000. Each village has an elected mayor or

village head. Health care is provided hierarchically from the provincial hospital to the district hospitals to the village-based public health offices.

Nearly all sub-districts (tumbons) have a government school. Like the political and health care system, the educational system is organized hierarchically from the center of the province to the districts and to sub-districts and villages. Periodic meetings of school administrators are held to disseminate important information. The structure and efficiency of the school system is conducive to the study of diseases that primarily afflict school-aged children.

Partners in the collaboration contributed from their strengths, and the convergence of capabilities facilitated the many successful investigations that were carried out. KPP contributed medical facilities, understanding of the impact and management of local disease problems and their management, public health infrastructure, schools, and prospective volunteers. The Thai MOPH brought expertise in disease surveillance and outbreak control and perspective on disease priorities. Investigators from the US Army contributed advances in diagnosis, knowledge of vaccines, entomological support, and strategies for study design. Vaccine manufacturers provided vaccines. Investigators at universities in Thailand and the US contributed their skills as well.

This collaboration led to studies that contributed to regulatory approval of two vaccines (Japanese encephalitis and hepatitis A vaccines) in two countries (Thailand and the US), knowledge of the pathogenesis and epidemiology of dengue, and understanding of the transmission of influenza. The collaboration also provided models for two major vaccine (hepatitis E and human

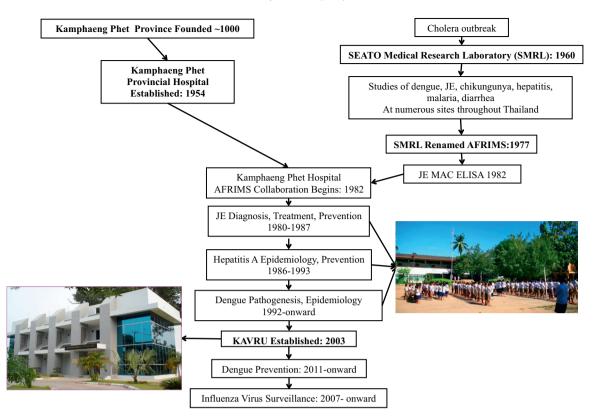


Fig. 1. Overview of collaborative studies performed in Kamphaeng Phet Province, Thailand.

immunodeficiency virus vaccines) trials conducted elsewhere. This paper reviews scientific progress derived from approximately 80 published studies carried out in KPP, emphasizing the personal and professional interactions that sustain such collaborations.

2. History of the collaboration

2.1. KPP Hospital

KPP Hospital was built in 1954 and expanded several times to the current 410 beds. The hospital cares for approximately 400,000 outpatients and 40,000 inpatients per year and has a modern laboratory and radiology department. Discharge records include listings of final diagnoses for all admitted patients. Because the hospital is the principal focus for medical care in the province, it is well suited to serve as the headquarters for population-based studies.

2.2. Thai Ministry of Public Health

In addition to many other missions, the Thai Ministry of Public Health conducted communicable disease surveillance that allowed public health leaders to identify a number of diseases, including Japanese encephalitis and dengue hemorrhagic fever, as substantial country-wide problems [2–5].

2.3. South East Asia Treaty Organization (SEATO) Medical Research Laboratory (SMRL), Bangkok, Thailand (Fig. 1) [1]

In 1958–59, in response to a cholera epidemic in Thailand, the Thai Government invited the US to join Thai investigators in understanding and managing the epidemic. A delegation, representing the US Navy, Army, and Public Health Service, visited Thailand's Undersecretary of State for Health, who had been a medical school roommate of a member of the US delegation. The King of Thailand

emphasized that the international research to be conducted by the new laboratory should benefit his subjects. The South East Asia Treaty Organization (SEATO) Thai Cholera Research Laboratory was thereby established, with the Undersecretary appointed the first Director General. In 1960, the Cholera Laboratory was renamed the SEATO Medical Research Laboratory (SMRL), with Thai and US components (Fig. 1).

2.4. The Armed Forces Research Institute of Medical Sciences (AFRIMS)

In 1977, SEATO was disbanded due to waning political interest in the overall organization, and the SMRL was renamed the Armed Forces Research Institute of Medical Sciences (AFRIMS), which is led by a Thai Military Director General. The U.S. Army component is commanded by a US Army Officer who reports to the Walter Reed Army Institute of Research (WRAIR) Commander. When US Component studies involve human subjects, the study protocols must be approved by both the Thai and US Army Institutional Review Boards.

In the years since the formation of AFRIMS, and prior to the collaboration described in this paper, AFRIMS scientists and Thai colleagues had made notable contributions in several areas of mutual interest, including dengue (pathogenesis [6], neutralizing antibody [7,8], epidemiology [9], and surveillance [10]), chikungunya [11], Japanese encephalitis in northern Thailand [12], malaria treatment [13], vectors of dengue, malaria, and Japanese encephalitis [14], and hepatitis B antigenic subtypes [15]. These studies had been conducted in a number of sites throughout Thailand, and with the experience gained from these prior collaborations, coupled with surveillance of infectious disease health threats by scientists in the Thai Ministry of Public Health, teams from AFRIMS and the Thai Ministry of Public Health were well-positioned to establish a new long term study site in Kamphaeng Phet, at which site methods

for control of some of these problems could be explored collaboratively.

2.5. Kamphaeng Phet-AFRIMS Virology Research Unit (KAVRU)

In 1982, AFRIMS scientists, seeking clinical specimens to evaluate a newly developed JE IgM antibody capture immunoassay (MAC ELISA) became aware that each summer the KPP Hospital admitted numerous pediatric patients with encephalitis. The hospital pediatric discharge diagnosis logbook revealed that the hospital provided care for many patients with encephalitis, as well as to patients diagnosed as having hemorrhagic fever, hepatitis, diarrhea, and pneumonia, suggesting a blueprint for multiple collaborative studies in the future.

As these studies unfolded, the hospital provided laboratory facilities and office space while AFRIMS provided technical support for virological diagnosis. The JE MAC ELISA evaluation was the first of many studies that led AFRIMS staff to become integrated with the community (doctors, nurses, school administrators, teachers, and parents) in the joint pursuit of public health research.

By 2001 the virology research laboratory was recognized as a valuable addition to the hospital's capabilities. Expansion was needed. Land was offered on the campus and funds for a new laboratory were obtained. The new laboratory was built and opened in 2003 as the KPP-AFRIMS Virology Research Unit, or KAVRU. The opening of KAVRU was co-officiated by the Permanent Secretary of the Thai Ministry of Public Health and the U.S. Ambassador to Thailand, as a joint Thailand-U.S. laboratory for the study of dengue and other emerging diseases. KAVRU was constructed as a state-of-the-art facility with its first floor dedicated to the clinical evaluation of subjects and processing/storage of specimens and the second floor to laboratory evaluation (including polymerase chain reaction testing). In light of increasing international requirements for vaccine trials, greater resources were applied to preparing the KAVRU study teams to comply with the latest regulatory standards.

3. Scientific results of the AFRIMS-KPP collaboration (Table 1)

3.1. Japanese encephalitis (Fig. 2)

In past years, AFRIMS investigators had investigated the ecology and epidemiology of JE in Chiang Mai province, and these studies laid the foundation for more sustained efforts in Kamphaeng Phet [21]. MOPH investigators conducted increasingly detailed surveil-lance of encephalitis [2] throughout Thailand, providing the MOPH of detailed knowledge of provinces with greatest risk (Table 1 and Fig. 2).

3.1.1. JE IgM antibody capture ELISA

The JE MAC ELISA test [16,17,19] that catalyzed the collaboration also revolutionized the diagnosis of arboviral encephalitis. Previously, etiological diagnosis had required a labor-intensive assay, performed on acute and convalescent sera, which provided a result long after discharge [21]. The new test could be performed quickly on small volumes of serum or CSF. The assay was used to establish the kinetics of antibody formation [18] and evaluate factors associated with fatal outcome [20]. The lead investigator shared data obtained in KPP with the US Centers for Disease Control Division of Vector Borne Diseases in Fort Collins, Colorado. AFRIMS offered training programs on performing the assay, eventually training scientists from many countries and serving as a reference laboratory as kits became commercialized [22–25]. The assay also facilitated performance of therapeutic studies. For example, dexamethasone had been used in treating JEV encephalitis patients, but a

Table 1

Results of studies of Japanese encephalitis, hepatitis A, dengue, and influenza conducted in Kamphaeng Phet Province.

```
JE diagnostic test (14 papers published)
```

Anti-JE IgM antibody detected in cerebrospinal fluid [16] and serum [17] by day 3 [18]

Many CSF specimens could be processed, facilitating disease surveillance [19]

Fatal outcomes were correlated with CSF virus, low antibody levels, and coma [20]

AFRIMS serves as reference laboratory for JE diagnostic kits [22–25] Use of dexamethasone in treatment of acute JE does not increase survival [26]

JE vaccine (1 paper published)

JE vaccine was safe in Thai children with an efficacy of 91% [27] The placebo cohort provided a prospective estimate of JE incidence [31] Following an additional study [28], US FDA approved BIKEN JE vaccine JE vaccine in Thailand introduced as routine immunization in Thailand, reducing rates

In the US, a cell culture JE vaccine, invented at WRAIR, licensed on basis of immunogenicity non-inferiority to vaccine tested in KPP [33,34] KPP experience contributed to recommendations for global control of JE [35,36]

Hepatitis A (5 papers published)

Enrolled children from KPP: 65,000 for surveillance, 40,000 for vaccine trial

Hepatitis A incidence in KPP estimated to be about 11/100/year [47] Hepatitis A vaccine was safe and 94% efficacious [49]

Licensure of the vaccine in the US was based in part on KPP trial results [50]

HAV vaccine was judged as not cost effective in Thailand [51] Following use of HAV vaccine in the US Military, the hepatitis A incidence fell [53]

Dengue (56 papers published)

In JE vaccine study, DHF affected 107/100,000 and dengue fever, 135/100.000 [27]

Dengue and JE MAC ELISA calibrated to distinguish between dengue and JE [32]

Severity was correlated with higher viremia, antibody response, and DEN 2 [56,57]

Certain HLA types are at risk of more severe illness [58]

Rare neurological manifestations of dengue were identified [107]

Antibody titers may not be good markers of immunity [86]

The focal nature of dengue transmission was illuminated [80,93,103,113] Site was prepared for phases 2 and 3 studies of dengue vaccines in rural Thai setting

17,815 children were enrolled in various studies Phase 3 trial of dengue vaccine is ongoing

Influenza (4 papers published)

Participated in global influenza surveillance efforts [116–119] An 800 person cohort was established for monitoring exposure to influenza viruses

Avian influenza associated with gender, smoking, and absent indoor plumbing [117]

placebo-controlled study revealed no improvement in case fatality ratios in such patients [26].

3.1.2. JE vaccine in KPP

In 1982, investigators at AFRIMS asked the Director of the Thai Center for Disease Control about possible collaborative work. The Director presented surveillance data that revealed sharp annual outbreaks of encephalitis in northern provinces, making these outbreaks possibly amenable to control through the use of a vaccine if one was available [2]. JEV vaccines had been used in Japan, but they had not been shown to be efficacious in Thailand. The president of a company that made JE vaccine who was also associated with the Japan International Cooperation Agency (JICA) agreed that his company would provide vaccine and placebo for an efficacy trial. The head of the Thai National Institutes of Health had long felt that an efficacy trial of a JE vaccine in Thailand was needed, and she offered

Past studies of JE epidemiology and ecology, relied on hemagglutination inhibition tests often done on fractionated sera. Improved diagnosis was needed JE Diagnosis improved: JE MAC ELISA Surveillance for JE and other arboviruses IgG and IgM Antibody In CSF and Serum improved, especially in Nepal IgG and IgM Antibody Kinetics Health Department Laboratories Trained JE Treatment: Dexamethsone does not reduce mortality AFRIMS serves as Reference Lab comparing assays Immunogenicity demonstrated .IE Vaccine (mouse brain) Thai FDA Approval in US Troops Efficacions JE Vaccine manufactured in Thailand Data used to support Thai effort to include Cell culture inactivated JE vaccine JE vaccine in EPI invented at WRAIR, FDA approved US FDA Approval based on non-inferiority to mouse JE Rates Fall brain vaccine Available for Global burden of JE estimated Thailand Switches to SA-14-14-2 Travelers. using placebo rate live JEV Vaccine including children

Fig. 2. Japanese encephalitis studies performed in Kamphaeng Phet Province by Thai Ministry of Public Health and US Army and the impact of those studies on Japanese encephalitis in Thai children and in US travelers.

her support. The provincial hospital director in KPP confirmed his support of an efficacy study.

Discussions between AFRIMS representatives and the Deputy Prime Minister of Thailand, members of Parliament, the Ministers of Health and Education, and the provincial governor, village heads, directors of primary and secondary schools, teachers, and parents provided a means by which all could be informed by the investigators. A protocol for the trial had been drafted, and the protocol was reviewed and approved by the Human Subjects Research Review Board of the US Army Surgeon General and the Thai MOPH Institutional Review Board, which in turn appointed a Thai oversight board to monitor the trial.

Between December 1984 and April 1985, more than 60,000 children from 496 villages in KPP were enrolled with parental consent and were vaccinated with one of two inactivated JE vaccines or tetanus toxoid [27]. The JE vaccine was made of virus purified from the brains of mice that had been inoculated with JE virus and inactivated with formalin. Enrollment and initial vaccination was accomplished by six teams, each team visiting three villages per day, followed by a second visit a week later to give a second dose. Each of the teams was led by a physician, and the MOPH provided many of these physicians from the Field Epidemiology Training Program conducted by the US Centers for Disease Control in Thailand. AFRIMS and KPP Hospital provided three nurses per team.

The JE vaccine efficacy was 91% [27], and these data contributed to Thai and US FDA licensure. The US FDA required one additional study to confirm safety and immunogenicity [28]. The Japanese company partnered with a company in the US to sponsor a New Drug Application and distribute vaccine. In Thailand the company facilitated construction of JE vaccine manufacturing facility. JE vaccine was phased into the Expanded Program on Immunization (EPI) throughout Thailand, resulting in a substantial decline in reported encephalitis cases [29,30]. Later, data from the placebo group provided rates of JE, contributing to global estimates of the burden of JEV [31].

In Thailand, a live attenuated JE vaccine has recently replaced the inactivated mouse brain vaccine, while, in the US, an inactivated JE vaccine produced in cell culture was invented at WRAIR as a replacement for the mouse brain vaccine tested in KPP, developed, under license to the Army, by a commercial manufacturer, and approved by the US FDA based on demonstration that the new vaccine was not inferior, with respect to stimulation of neutralizing antibody, to the vaccine that had been tested in KPP [33,34]. In addition, expertise gained in KPP contributed to recommendations for global control of JE [35,36].

Other studies demonstrated the presence of JE antigen in neurons in fatal cases [37], that non-immune pigs could be effective sentinels for JE virus circulation [38], that JE antibodies were detected in cells, blood and cerebrospinal fluid [39], and that JEV disease was more severe in patients in whom encephalitis preceded an immune response [42].

3.1.3. Entomological studies

Entomological studies revealed that mosquito cells derived from *Aedes pseudoscutellaris* were useful for bedside inoculation of specimens: serum and CSF from non-fatal JE cases failed to yield virus, while virus could be cultured from CSF from fatal cases or fresh brain collected at autopsy [40]. Isolations of virus from pooled collections of mosquitoes revealed that JE virus could be isolated for only a 10-day period around the time when the peak incidence of disease was observed in humans [41], suggesting that natural factors supporting virus transmission were transient.

3.2. Hepatitis A studies (Fig. 3)

3.2.1. Invention of hepatitis A vaccine

During the time in which the trial of JE vaccine was conducted in KPP, scientists at WRAIR had invented a test for detecting neutralizing antibody against hepatitis A [43], invented the world's first inactivated hepatitis A vaccine (HAV) [44], and protected monkeys against challenge. A clinical trial showed that the vaccine

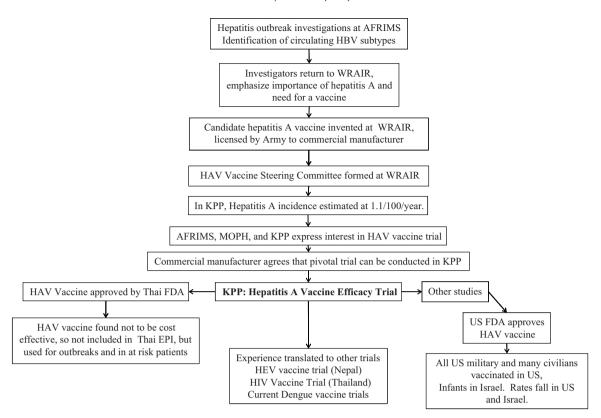


Fig. 3. Hepatitis A studies performed in US and Thailand, particularly in Kamphaeng Phet Province, leading to approved HAV vaccine.

stimulated neutralizing antibody in all eight volunteer recipients [45]. Using a "No Dollar Agreement", the Army transferred the HAV vaccine technology to a commercial vaccine manufacturer who in turn became the regulatory sponsor of the vaccine [46] (Fig. 3).

3.2.2. Hepatitis A vaccine trial in KPP

The manufacturer initially favored an efficacy trial in the US, but negotiations for such trials were unsuccessful, while epidemiological studies in KPP suggested that rates of hepatitis A (1.1 cases/100 children/year) [47] were substantial. Investigators at AFRIMS, along with Thai medical and public health leaders, felt that a collaborative HAV vaccine efficacy trial, similar to the recent JE vaccine efficacy trial, but with improvements in design, could be conducted in KPP. Eventually the company was convinced, and a collaborative team was assembled to conduct the second large-scale vaccine trial in Thailand. Approximately 42,000 children were enrolled to receive either hepatitis A or B vaccine, followed eventually by a crossover immunization. Surveillance for hepatitis A cases in the study population was conducted using a case definition of missed school, liver enzymes elevated above the upper limit of normal, and a positive test for hepatitis A IgM. Although not initially a part of the case definition, the US FDA requested that cases be confirmed by RT-PCR detection of viral RNA in feces, and, following substantial effort, data from this testing was added to the license application. Study participants ultimately received both hepatitis A and B vaccines and were provided medical care, and communities benefited by mitigation of hepatitis A outbreaks through construction of wells and renovation of toilet and hand washing facilities in schools.

The hepatitis A vaccine was found to be safe and 94% efficacious [48,49]. Publication of the results with Thai and US Army authors attested to the breadth of the collaboration. The vaccine was licensed in the US and Thailand based on data from KPP [50]

and other studies, several of which were performed by Army investigators in military populations [46].

3.2.3. Post trial impacts

Analyses suggested that universal childhood immunization against HAV would not be cost effective in Thailand [51], so the vaccine was not incorporated into the routine immunization program. However, the HAV vaccine is used in Thailand for control of outbreaks, post exposure prophylaxis, and protection of persons at high risk of exposures. Lack of universal use of the HAV in Thailand provided the opportunity to discuss the ethics of conducting a trial after which vaccine was made available to populations other than those that participated in the trial [52].

In the United States, use of HAV vaccine was associated with a dramatic reduction of numbers of cases, especially in the military [53]. An even more precipitous decline in cases in all ages was observed in Israel following widespread immunization of toddlers [54].

3.3. Dengue

AFRIMS investigators and Thai colleagues had contributed to the understanding of the pathogenesis of dengue hemorrhagic fever [6]. Studies had relied on traditional diagnostic tests, including hemagglutination inhibition and neutralizing antibody assays [7,8] and various means of isolating dengue virus from clinical specimens, including mosquito inoculation and testing of tissues with immunofluorescence. A major advance had occurred with the discovery at WRAIR and distribution by the US CDC of mouse monoclonal antibodies specific for serotypes of dengue and other flaviviruses [56–58]. This monoclonal antibody technology was adapted rapidly at AFRIMS and expanded. Thai clinicians, particularly at Bangkok Children's Hospital, were recognized as experts in the diagnosis and clinical management of dengue hemorrhagic

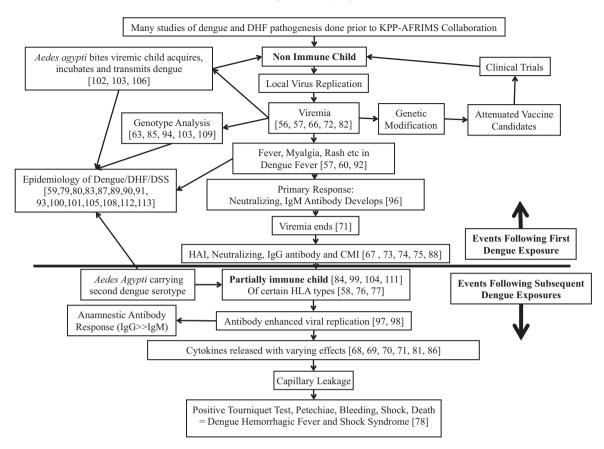


Fig. 4. Recent studies of dengue transmission, pathogenesis, diagnosis, and genetics conducted in Kamphaeng Phet Province.

fever, with diagnostic support provided by nearby AFRIMS. With the advent of improved techniques for detection of cytokine mediators in the immune response to dengue, investigators were in excellent position to conduct detailed studies at a fixed facility in KPP.

3.3.1. Dengue diagnosis

For many years serological diagnosis of dengue had relied on the hemagglutination inhibition test, which was challenging because of the great amount of cross-reactive antibody detected in this assay following any flavivirus infection. Surveillance in the JE vaccine efficacy trial revealed that dengue hemorrhagic fever hospitalizations occurred frequently, indicating that dengue was a significant problem for the children of KPP, providing an opportunity to refine the ELISA test to distinguish JE and dengue infections serologically in an area where both circulate [32] and suggesting that dengue vaccines might be evaluated in future studies. Analysis of specimens collected in KPP allowed refinement of the MAC ELISA assay used for JEV infections to diagnose JEV and dengue virus infections in an area where both viruses co-circulate [27].

3.3.2. Dengue pathogenesis (Fig. 4)

Dengue virus is a major cause of morbidity in Thailand, and Thai physicians have been leaders in understanding dengue pathophysiology, classification of cases, clinical management, and surveillance [55]. AFRIMS supported presentations in the diagnosis and management of dengue by leading Thai authorities to the staff at KPP Hospital (Fig. 4).

In 1994, investigators from the University of Massachusetts, Yale Arbovirus Research Laboratory in the US and, in Thailand, Siriraj Medical Molecular Biology Center, the Siriraj Hospital Institute of Pathology and Department of Transfusion Medicine, and the MOPH,

began participation in studies that spanned 15 years, conducted largely at KPP Hospital, to better define the pathophysiology of dengue disease, provide insights into methods of prevention and treatment, and prospectively evaluate dengue virus transmission and disease in primary school children. More severe dengue was associated with higher viremia titers, secondary immune response, and dengue type 2 infection [56,57], and certain human leukocyte antigen types appeared to be at risk of more severe disease [58]. The burden of dengue disease appeared to be greater than estimated from surveillance data [59]. Additional studies have yielded a great deal of information about dengue in Thailand [60–113].

3.3.3. Dengue vaccine trials

Several dengue vaccine candidates were under development in different laboratories. By 2010, the manufacturer of a candidate dengue vaccine began preparations for a global multi-country Phase 3 trial, and KPP was identified as one of two sites in Thailand to be included. In collaboration with KPP Provincial Hospital, the Thai MOPH, Faculty of Tropical Medicine Mahidol University, the US Army, and the manufacturer, a Phase 3 dengue vaccine trial was initiated in Thailand in 2011. The manufacturer adopted surveillance methods developed in KPP for application to the entire phase 3 program. Vaccinations in the trial were completed in 2012 and active surveillance for dengue virus infections is ongoing.

3.3.4. Dengue vectors

Standardization of container descriptions and mosquito collection methods may facilitate evaluations of control strategies [90,108]. Dengue virus adaption to *Aedes agypti* populations depends on both virus and vector genotype [95]. Studies of breeding sites of *Aedes agypti* may lead to improved methods of vector control programs [100], as may observations that some

households contribute disproportionately to dengue transmission [103], although residents may take measures against vectors mainly in response to the perceived nuisance of mosquitos [89].

3.4. Respiratory and other disease studies

Samples collected in the dengue studies were used in understanding the presence of other pathogens, including leptospirosis and metapneumovirus [110,114,115].

Prospective respiratory disease surveillance began in 2007, and KAVRU participated as a global influenza surveillance site [116–119]. With cases of H5N1 detected in both birds and humans in KPP, the need for influenza testing as a service to the province became apparent. In 2008, KAVRU began testing samples from the hospital and established PCR testing of respiratory samples on site. Subsequently, a cohort was enrolled to better understand human exposure to zoonotic influenza in KPP which suggested that exposure to avian influenza virus might be more common than previously thought [117].

4. Discussion

The establishment of a study site to evaluate a diagnostic test for JEV infection led, over 30 years, to studies supporting licensure of two vaccines, substantial scientific accomplishments, surveillance of etiologically confirmed disease, and pivotal experiences for scores of Thai and US researchers.

4.1. Site selection

Initially, the collaboration resulted from a simple site selection effort, guided by the documented presence of diseases of mutual interest and by personal relationships. Established Thai scientists at AFRIMS provided an essential network of expertise and trust among Thai governmental authorities. Established on a foundation of mutual interests, the collaboration grew over the years and included the construction of a modern laboratory (KAVRU) to provide laboratory support to ongoing studies of endemic and emerging diseases.

4.2. Beneficial joint collaboration

A hallmark of this collaboration has been the sensitive interaction by all participants to assure that studies addressed the expectations of sponsoring agencies. The Thai MOPH guided the collaboration toward the needs of the Thai people, as had been requested during the founding of the SMRL. Medical and educational authorities in KPP retained the focus on diagnosis, treatment and prevention of diseases affecting the people. AFRIMS investigators assured that studies addressed the needs of the US Army, which furnished resources programmed to find means to protect military personnel. Vaccine manufacturers contributed vaccine technology. Authorship on publications resulting from this collaboration reflects the participation of these agencies.

4.2.1. Shared experience

Both the Thai and US investigators grew through the shared experience derived from the many studies conducted in KPP. AFRIMS Department of Virology leadership rotated every 4–8 years, providing new perspectives, a link to technology under development at WRAIR, and experience with study design. The senior Thai investigators in the department provided continuity, expertise in diagnostic assays, and, most important, an interface with the Thai medical establishment. KPP hospital directors provided

stability as well and participated in selection of studies and ongoing problem solving. Thai physicians and nurses possessed significant expertise in the treatment of these diseases they encountered on a daily basis. Thai MOPH leaders conducted careful disease surveillance and were well aware of epidemiological problems and interventions in need of study. Pooling of these talents resulted in research teams able to approach large public health problems.

A highly purified JEV vaccine, developed initially in the US during World War II to protect American Servicemen [120] and in Japan following World War II for the benefit of Japanese children, was evaluated in KPP, and the results of this evaluation led to the introduction and manufacturing of this vaccine in Thailand for the benefit of Thai children and to the licensure by the US FDA of the vaccine for the benefit of military personnel and travelers to JE endemic areas, completing an interrupted 40 year long process

The HAV vaccine, invented at WRAIR, was also shown to be efficacious in KPP, providing additional experience for US and Thai investigators. A dengue vaccine trial is underway, and testing of additional dengue vaccines candidates is expected.

4.2.2. Applicability of scientific results in KPP to development of vaccines against a range of tropical infectious diseases

Studies in KPP that established the efficacy of the mouse brain derived JEV vaccine and of the HAV vaccine had direct bearing on more recent efforts to develop vaccines against these diseases. For example, now at WRAIR, the leader of the HAV vaccine trial assembled a group that invented an inactivated cell culture JEV vaccine produced in vero cells that became licensed in the US based on demonstration of serological non-inferiority to the mouse brain vaccine tested in Thailand, with no additional efficacy demonstration required

The HAV vaccine study had a direct bearing on a study of the efficacy of a baculovirus-expressed recombinant HEV vaccine by reinforcing the need for a case definition that included clear evidence of liver injury, virus detection by PCR, multiple specimens collected early in disease, and longer follow up of cases. Conducted in 2000 Nepalese soldiers by an international team of investigators at another AFRIMS laboratory, known as the Walter Reed Army Research Unit-Nepal (WARUN), the study demonstrated that the efficacy of the HEV vaccine was 95% [121,122]. Although the hepatitis E vaccine proved to be highly efficacious, the manufacturer of this particular candidate has not further developed it, in part because of uncertainties regarding the public health priority for hepatitis E prevention and control through immunization.

The trials of JEV and HAV vaccines together set examples for the large-scale HIV vaccine efficacy trial conducted later as a Thai-US collaborative effort [123]. Expanded dengue vaccine trials, now being conducted in KPP, will benefit from the experiences obtained in earlier trials in that province.

4.2.3. Medical benefits

As a result of studies conducted in KPP, a number of specific products became available for much larger populations. In Thailand, studies suggested that the JEV vaccine would be cost effective [124,125] and vaccine usage was systematically expanded to progressively greater numbers of provinces, followed by a decline in cases reported, suggesting that thousands of cases have been prevented by implementation of JE immunization.

4.2.4. Community benefits

Nearly 200,000 residents of KPP volunteered for the studies that have been conducted in their province. Informal follow up

Table 2Challenges in conducting field studies and the lessons learned from the m.

Positive/negative aspects of trials	Lesson learned
Japanese encephalitis efficacy trial	
Initially, relationships between agencies (AFRIMS, MOPH, KPP Hospital, and BIKEN) were	Communication by investigator team and concerned parties may
poorly defined. Investigator provided all parties with a weekly written update	help cement relationships
Prior to ICH Guidelines, investigators improvised many solutions	ICH guidelines provide useful checklists for investigators
A serious adverse event in a study participant led investigators to consider terminating the	An oversight board, sometimes called a Data and Safety
study. Following investigation, the MOPH Oversight Board recommended that the trial continue	Monitoring Board, can provide important guidance
Efficacy trial results were published, but impact in Thailand required manufacture and	"Excellence in Research is Not Enough": Research must be followed
introduction of the vaccine and in the US, the FDA required one additional study for licensure	by regulatory and logistical activities and vaccine distribution
Hepatitis A	
Invention and testing of the vaccine required interactions of WRAIR, NIAID, AFRIMS, MOPH,	Relationships are essential elements of success in complex
KPP, and a manufacturer	international endeavors
Based on unfavorable cost benefit studies, HAV vaccine was not introduced into the Thai EPI, though it was used for outbreak suppression and other focal uses	Re-analysis of costs and benefits may suggest suitable remedies
US FDA unexpectedly requested demonstration of etiologic agent in stool specimens from	Robust case definitions, with demonstration of etiological agents,
cases	should be agreed to at outset
Dengue	
Quality control of clinical observations, such as temperature, blood pressure, spleen size	Care must be exercised that all participating staff are trained to
determinations, presence of petechia, and of serological testing may require	obtain data in a uniform and reproducible way and that all
standardization, training, and validation as well as photographic documentation. Methods	measurements are validated before beginning phase three trials
for "routine tests" like virus neutralization tests, may vary greatly from lab to lab	
Many competing vaccines, rising cost of studies, and commercial interests that may differ	Strive for common diagnostic tests and comparative (head to
from public health considerations complicate the selection of candidate vaccines and sites	head) trials with predefined endpoints whenever possible
Influenza	
Adding new tests and procedures presented challenges in standardization and quality	Special training for all procedures should be sought at global
control	centers of excellence

conversations, provided by one of the authors (BLI), suggest that the organized clinical trials, with the informed consent processes that were mandated by the Institutional Review Boards resulted in a better-informed and medically aware citizenry in the province [126].

4.2.5. Challenges and positive/negative aspects of the conduct of the trials and lessons learned from conducting these studies (Table 2)

Studies that involve scores of investigators from different countries, multiple governmental Institutional Review Boards (IRBs), and hundreds of thousands of subjects are filled with complexity. Millions of data points may converge to answer a single question: Is the vaccine efficacious or not? With such complexity, there is room for learning many lessons, many of which are now incorporated into the International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use (ICH) guidelines for Good Clinical Practices, a document that was not available in 1982 [127] (Table 2).

In addition, several overarching principals are related to the conduct of studies with investigators of different nations, governed by different rules, working in different languages with parents and subjects who recognize the medical problem from its effects on their community. To identify research needs, potential investigators must be attuned to the advice of mentors and consultants. Where multiple vaccine candidates exist, head-to-head trials are desirable, but often resisted by manufacturers, so standardization and validation of case definitions and serological tests may suffice to facilitate informative comparisons.

Second, complex studies require a great deal of trust between individuals and between organizations, which must ultimately work as a team. Teams are said to move through several stages, including "Forming, Storming, Norming, and Performing" as they mature [128]. Bringing together IRB reviewers, oversight committees, vaccine manufacturers, study populations, collaborating investigators, and scores of supporting staff members is, in effect,

an exercise in team building. Teams working in KPP followed this pattern, initially finding their way through occasionally turbulent waters, eventually developing trusting relationships, with many years of performing successful studies. The founding of KAVRU represents a physical manifestation of team building over the decades of collaboration before and after its founding.

Third, the subtle details related to the collection of clinical observations, and analysis of specimens must be carefully managed to prevent unintentional degradation of study results. Careful training and validation of all measurements and assays is essential.

Finally, elegant study design and compelling results are of little worth if the findings do not result in meaningful implementation. Following the JEV vaccine and HAV vaccine efficacy trials, both vaccines were advanced substantially toward licensure, and the results of the two studies carried out in KPP are mentioned prominently in the package inserts. Both vaccines became available for use in US citizens. HAV cases have fallen dramatically in the US since the introduction of the vaccine. In Thailand, the MOPH determined that the JEV vaccine would be cost effective, manufacturing facilities were constructed, and the vaccine added to routine immunizations with a reduction in the numbers of encephalitis cases. The hepatitis A vaccine was not introduced as a routine immunization in Thailand, a result that may have been related to the relative mildness of hepatitis A infections in young people. Perhaps conclusions will be altered if hepatitis surveillance should find a growing burden of disease of if vaccine cost should

4.3. Future studies

The success of the studies of JEV and HAV and the initiation of the ongoing dengue vaccine study, combined with the characterization of the province's disease burden, suggest that additional future studies, perhaps of dengue and influenza, done at KAVRU may yield valuable information and products for treatment or prevention of these illnesses.

Table 3

Acknowledgment of contributions.

Acknowledgment of contributions.	
Participant	Role
Albert Sabin	Led effort to make JEV Vaccine during WWII
Richard Mason	Member of founding delegation from WRAIR
Kenneth Goodner	Provided introduction to Thai Classmate from Jefferson Medical College
Luang Binbakya Bidybhed	As Thai Undersecretary of State for Health, arranged initial meeting with King of Thailand. First Director General of
	SEATO Cholera Research Laboratory
Franklin Top	Established filter paper method of transporting dried sera to lab for testing for dengue antibody
Phillip Russell	As investigator at AFRIMS, co-developed dengue neutralizing antibody assay. As Commander, USAMRMC, supported
	concept of Japanese encephalitis vaccine efficacy trial
William Bancroft	Conducted studies of hepatitis A and dengue at AFRIMS and provided leadership for HAVV at WRAIR
Robert McNair Scott	Performed epidemiological studies of dengue. Led HEVV study in Nepal
Michael Benenson, Frank Sodetz	Commanders, USA Medical Component, AFRIMS during JEV studies
Ananda Nisalak	AFRIMS' senior Thai investigator, known and respected throughout Thailand. Co-Inventor of JE MAC ELISA test.
	Suggested test site in KPP. Coordinated large JE vaccine efficacy trial. Provided
Donald Burke	Invented test for Japanese encephalitis and supported JEVV trial
	Launched (with Dr Ananda) the first AFRIMS studies in KPP
	Oversaw invention of inactivated hepatitis A vaccine
Michael Ussery	AFRIMS co-investigator of test for Japanese encephalitis
Thanom Laorakpongse	KPP Hospital Director, senior collaborator
Nursing and Lab Staff	At KPP Hospital Provided patient care and participated in studies
Nursing and Lab Staff	At AFRIMS, provided clinical data collection for clinical and epidemiological studies and vaccine trials. Provided
	laboratory test development and support
Charles Hoke	At AFRIMS, formulated and led JE vaccine efficacy trial and dexamethasone treatment study, at WRAIR, chaired
	hepatitis and flavivirus vaccine steering committees and at USAMRMC, directed Military Infectious Diseases Research
	Program
Somnuek Lamjiak	Provided nursing support, unacknowledged in JEV vaccine trial paper
Suchard Jetanesen	At Thai MOPH CDC, conducted disease surveillance for Thailand that identified location of annual encephalitis
	outbreaks
Natth Bhamaraphavati	Distinguished Thai physician/scientist who suggested link between AFRIMS and Thai NIH
Nathirat Sangawhipa	At Thai NIH, identified need for JE Vaccine efficacy trial and supported proposed trial
Konosuke Fukai	At BIKEN Foundation, manufactured JEVV used in efficacy trial
Walter Brandt	At USAMMDA, convened meeting at USFDA, including vaccine manufacturer and US distributor, to review JEVV
W	efficacy data and discuss additional data required for licensure in the US
Mary Kaye Gentry, Erik Henchal,	At WRAIR, Identified and evaluated monoclonal antibodies for identifying and typing dengue and Japanese
Walter Brandt, Joel Dalrymple	encephalitis and other flaviviruses
Edmund Tramont	Supported protocol for JE Vaccine Efficacy Trial
Stanley Lemon	Developed tests and laid groundwork for HAVV development
Leonard Binn	At WRAIR, invented first HAVV and showed protection of monkeys. Developed neutralizing antibody test
Kenneth Eckels	At WRAIR, manufactured first GMP batch of hepatitis A vaccine
Maria Sjogren John Boslego	Led first clinical trial of WRAIR's hepatitis A vaccine
Bruce Innis	AFRIMS commander during hepatitis A vaccine efficacy trial At AFRIMS, supported JEVV trial, advanced study of ELISA test for JE and dengue, conducted studies of HAV, proposed
bluce lillis	and led HAVV trial, proposed hepatitis E vaccine efficacy trial in Nepal, and organized team at WRAIR to invent
	replacement JEV vaccine produced in vero cells
Rapin Snitbahn	AFRIMS' senior Thai hepatitis investigator
Prayura Kunasol	As head of Thai CDC, advocated for HAV vaccine trial
Erich D'Hondt	Coordinated HAVV technology transfer from WRAIR to SKB
David Krause	At SKB, coordinated clinical development of HAV vaccine
Suchitra Nimmannitya	Established grading of DHF and clinical treatment, Provided expert consultation on management of dengue to KPP
Siripen Kalayanarooj	hospital staff
Frank Ennis, Alan Rothman, Sharone Green	At University of Massachusetts, conducted numerous US NIH funded studies of dengue pathogenesis
Supamit Chunsuttiwat	Thai MOPH spokesperson for Thai Immunization Program that adopted JE vaccine. Co-investigator on dengue
Suparine Chansaterwae	pathogenesis studies
David Vaughn	Led numerous studies of dengue immunology and pathogenesis, validating procedures. Secured funding for KAVRU
Buvia vaugini	construction. Directed Military Infectious Diseases Research Program
Mammen Mammen	Led dengue school-based and village cluster studies in KPP and supported HEV vaccine trial in Nepal. Designed and
	executed the construction of KAVRU
James W. Jones	From AFRIMS and the University of California, Davis: Provided entomological studies following startup of KAVRU. Led
Tom W. Scott	the entomology contributions to KPSII cluster investigations
Timothy Endy	Conducted numerous school-based studies of dengue epidemiology in KPP and initiated the HEV trial in Nepal
Stephen Thomas	Studied dengue epidemiology and led WRAIR's dengue vaccine effort
Kamchai Rungsimunpaiboon	KPP Hospital Director
Tawee Chotpitayasunondh	As consultant, Thailand Ministry of Public Health, served as country coordinating investigator for dengue vaccine trial
Punnee Pitisuttithum	At Faculty of Tropical Medicine, Mahidol University, served as KPP Site Principal Investigator, dengue vaccine efficacy
	trial
Robert Gibbons	Led studies of dengue, Japanese encephalitis and influenza
	Conceived of the idea for this paper
In-Kyu Yoon	At AFRIMS, led virology department during dengue vaccine trials
Chusak Pimgate, Darunee Tannitisupawong	KAVRU site Directors since the opening of KAVRU
Chunlin Zhang	Established PCR capability at KAVRU in support of dengue cluster studies

Table 4
Acronyms used.

Name	Acronym
Armed Forces Research Institute of Medical Sciences	
Center for Disease Control	AFRIMS CDC
Cerebrospinal Fluid	CSF
Dengue Hemorrhagic Fever	DHF
Dengue Shock Syndrome	DSS
Enzyme Linked Immunosorbent Assay	ELISA
Expanded Program on Immunizations	EPI
Good Clinical Practices	GCP
Good Manufacturing Practices	GMP
GlaxoSmithKline	GSK
Hepatitis A virus	HAV
Hepatitis E virus	HEV
Human Immunodeficiency virus	HIV
Human Leukocyte Antigen	HLA
Immunoglobulin G	IgG
Immunoglobulin M	IgM
Immunoglobulin G antibody capture	GAC
Immunoglobulin M antibody capture	MAC
Institutional Review Board	IRB
International Conference on Harmonization of Technical	ICH
Requirements for the Registration of Pharmaceuticals for	
Human Use	
Japanese encephalitis virus	JEV
Japan International Cooperation Agency	JICA
Kamphaeng Phet Province	KPP
Kamphaeng Phet-AFRIMS Virology Research Unit	KAVRU
Ministry of Public Health	MOPH
National Institutes of Health	NIH
Reverse Transcriptase Polymerase Chain Reaction	RT-PCR
Ribonucleic Acid	RNA
SEATO Medical Research Laboratory	SMRL
SmithKline Beecham	SKB
South East Asia Treaty Organization	SEATO
United Nations Educational, Scientific, and Cultural Organization	UNESCO
United States Food and Drug Administration	USFDA
US Army Medical Materiel Development Activity	USAMMDA
US Army Medical Research and Materiel Command	USAMRMC
Walter Reed Army Institute of Research	WRAIR
Walter Reed Army Research Unit-Nepal	WARUN

5. Conclusions

The collaboration between the Thai MOPH, KPP, and AFRIMS has resulted in a productive relationship of 30 years, culminating in the establishment of the Kamphaeng Phet-AFRIMS Virology Research Unit (KAVRU) and in the publication of approximately 80 papers. There have been benefits to all partners: Participants in vaccine trials benefited directly by increased medical surveillance and/or by administration of a beneficial vaccine at the end of the trial. The children of Thailand benefited from the introduction of the JEV into the Expanded Program on Immunizations (EPI) and, to a lesser extent, from the targeted use of HAV vaccine. The local community received training and mentorship and improved facilities for diagnosis of IE, hepatitis, dengue, and influenza. Investigators participated in valuable studies of the efficacy of JEV vaccine and HAV vaccine, paving the way for ongoing studies of dengue vaccines and studies of HIV and HEV vaccines. The Thai MOPH furthered its mission by conducting encephalitis surveillance that illuminated a problem so that the need for vaccine could be perceived. The vaccine manufacturers received information that supported applications for regulatory approval and, in the case of IE, allowed Japan to provide assistance in improving Thai vaccine capabilities. The US Army was able to identify, test, and procure two vaccines to protect service members against longstanding threats to their health. The general population of the US obtained access to two products valuable to travelers. And, by demonstrating the efficacy of these two vaccines and the reduced incidence of both diseases following use of the vaccines, the global community received the benefit of a successful international collaboration contributing to the improvement of complex public health problems. The multitude of beneficial outcomes from basic research to applied public health is in keeping with the guidance provided by a former AFRIMS investigator, Commander of the WRAIR and of the Army Medical Research and Materiel Command and President of the American Society for Tropical Medicine and Hygiene that "Excellence in Research is Not Enough" [129] (Table 3).

The acronyms used in this study are listed in Table 4.

Funding

The US Army Medical Research and Materiel Command, the US National Institute of Allergy and Infectious Diseases, the Armed Forces Health Surveillance Center, Global Emerging Infections Surveillance and Response System, and the Thai MOPH provided funding for the studies described. GSK provided partial funding for the HAV efficacy trial. None of the funding agencies had any influence over the content of this paper. Sanofi Pasteur provided funding for the ongoing dengue vaccine trial.

Disclaimer

The opinions or assertions contained herein are the private views of the authors and are not to be construed as reflecting the official views of the United States Army or the United States Department of Defense.

Acknowledgements

Thanom Laorakpongse, now deceased, provided-ongoing support as the Director of KPP Hospital. The authors thank the people of KPP and the staff of KAVRU for their support of the numerous clinical trials mentioned in this paper.

Numerous individuals contributed to the design and execution of the studies described in this paper. A partial list of individuals and their contributions is included in Table 3.

References

- [1] Woodward TE. Introduction to the history of the Armed Forces Medical Units in Kuala Lumpur Malaya and the Armed Forces Research Institute of Medical Sciences (AFRIMS) in Bangkok; 2013. Thailand. 5 February 1987. Found at: http://www.afrims.org/weblib/eArchive/History%20of%20AFRIMS.pdf (accessed 23.02.13).
- [2] Gunakasem P, Chantrasri C, Simasathien P, Chaiyanun S, Jatanasen S, Pariyanonth A. Surveillance of Japanese encephalitis cases in Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 1981;12:333–7.
- [3] Gunakasem P, Chantrasri C, Chaiyanun S, Simasathien P, Jatanasen S, Sangpetchsong V. Surveillance of dengue hemorrhagic fever cases in Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 1981;12:338–43.
- [4] Jatanasen S. Occurrence of haemorrhagic fever in Thailand, 1958–64. Bulletin of the World Health Organization 1966;35:79–80.
- [5] Pant CP, Jatanasen S, Yasuno M. Prevalence of Aedes aegypti and Aedes albopictus and observations on the ecology of dengue haemorrhagic fever in several areas of Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 1973;4:113–21.
- [6] Halstead SB, Nimmannitya S, Cohen SN. Observations related to pathogenesis of dengue hemorrhagic fever. IV. Relation of disease severity to antibody response and virus recovered. Yale Journal of Biology and Medicine 1970;42:311–28.
- [7] Russell PK, Nisalak A. Dengue virus identification by the plaque reduction neutralization test. Journal of Immunology 1967;99:291–6.
- [8] Russell PK, Nisalak A, Sukhavachana P, Vivona S. A plaque reduction test for dengue virus neutralizing antibodies. Journal of Immunology 1967;99:285–90.
- [9] Russell PK, Yuill TM, Nisalak A, Udomsakdi S, Gould DJ, Winter PE. An insular outbreak of dengue hemorrhagic fever. II. Virologic and serologic studies. American Journal of Tropical Medicine and Hygiene 1968;17:600–8.

- [10] Top Jr FT, Gunakasem P, Chantrasri C, Supavadee J. Serologic diagnosis of Dengue haemorrhagic fever using filter paper discs and one Dengue antigen. Southeast Asian Journal of Tropical Medicine and Public Health 1975;6:18–24.
- [11] Nimmannitya S, Halstead SB, Cohen SN, Margiotta MR. Dengue and chikungunya virus infection in man in Thailand, 1962–1964. I. Observations on hospitalized patients with hemorrhagic fever. American Journal of Tropical Medicine and Hygiene 1969;18:954–71.
- [12] Grossman RA, Gould DJ, Smith TJ, Johnsen DO, Pantuwatana S. Study of Japanese encephalitis virus in Chiangmai Valley, Thailand. I. Introduction and study design. American Journal of Epidemiology 1973;98:111–20.
- [13] Pearlman EJ, Doberstyn EB, Sudsok S, Thiemanun W, Kennedy RS, Canfield CJ. Chemosuppressive field trials in Thailand. IV. The suppression of *Plasmodium falciparum* and *Plasmodium vivax* parasitemias by mefloquine (WR 142, 490, A 4-quinolinemethanol). American Journal of Tropical Medicine and Hygiene 1980:29:1131-7.
- [14] Gould DJ, Edelman R, Grossman RA, Nisalak A, Sullivan MF. Study of Japanese encephalitis virus in Chiangmai Valley, Thailand. IV. Vector studies. American Journal of Epidemiology 1974;100:49–56.
- [15] Snitbhan R, Scott RM, Bancroft WH, Top Jr FH, Chiewsilp D. Subtypes of hepatitis B surface antigen in Southeast Asia. Journal of Infectious Diseases 1975;131:708–11.
- [16] Burke DS, Nisalak A, Ussery MA. Antibody capture immunoassay detection of Japanese encephalitis virus immunoglobulin M and G antibodies in cerebrospinal fluid. Journal of Clinical Microbiology 1982;16:1034–42.
- [17] Burke DS, Nisalak A. Detection of Japanese encephalitis virus immunoglobulin M antibodies in serum by antibody capture radioimmunoassay. Journal of Clinical Microbiology 1982;15:353–61.
- [18] Burke DS, Nisalak A, Ussery MA, Laorakpongse T, Chantavibul S. Kinetics of IgM and IgG responses to Japanese encephalitis virus in human serum and cerebrospinal fluid. Journal of Infectious Diseases 1985;151:1093–9.
- [19] Burke DS, Nisalak A, Hoke Jr CH. Field trial of a Japanese encephalitis diagnostic kit. Journal of Medical Virology 1986;18:41–9.
- [20] Burke DS, Lorsomrudee W, Leake CJ, Hoke CH, Nisalak A, Chongswasdi V, et al. Fatal outcome in Japanese encephalitis. American Journal of Tropical Medicine and Hygiene 1985;34:1203–10.
- [21] Grossman RA, Edelman R, Willhight M, Pantuwatana S, Udomsakdi S. Study of Japanese encephalitis virus in Chiangmai Valley, Thailand. 3. Human seroepidemiology and inapparent infections. American Journal of Epidemiology 1973;98:133-49.
- [22] Jacobson JA, Hills SL, Winkler JL, Mammen M, Thaisomboonsuk B, Marfin AA, Gibbons RV. Evaluation of three immunoglobulin M antibody capture enzyme-linked immunosorbent assays for diagnosis of Japanese encephalitis. American Journal of Tropical Medicine and Hygiene 2007;77:164–8.
- [23] Wierzba TF, Ghimire P, Malla S, Banerjee MK, Shrestha S, Mammen MP, et al. Laboratory-based Japanese encephalitis surveillance in Nepal and the implications of these results for a national immunization strategy. American Journal of Tropical Medicine and Hygiene 2008;78:1002–6.
- [24] Khalakdina A, Shrestha SK, Malla S, Hills S, Thaisomboonsuk B, Shrestha B, et al. Field evaluation of commercial MAC ELISA diagnostic tests for the detection of Japanese encephalitis virus infection among encephalitis patients in Nepal. International Journal of Infectious Diseases 2010;14(S3):e79–84.
- [25] Moore CE, Blacksell SD, Taojaikong T, Jarman RG, Gibbons RV, Lee SJ, et al. A prospective assessment of the accuracy of commercial IgM ELISAs in diagnosis of Japanese encephalitis virus infections in patients with suspected central nervous system infections in Laos. American Journal of Tropical Medicine and Hygiene 2012;87:171–8.
- [26] Hoke Jr CH, Vaughn DW, Nisalak A, Intralawan P, Poolsuppasit S, Jongsawas V, et al. Effect of high-dose dexamethasone on the outcome of acute encephalitis due to Japanese encephalitis virus. Journal of Infectious Diseases 1992;165:631–7.
- [27] Hoke CH, Nisalak A, Sangawhipa N, Jatanasen S, Laorakapongse T, Innis BL, et al. Protection against Japanese encephalitis by inactivated vaccines. New England Journal of Medicine 1988;319:608–14.
- [28] Defraites RF, Gambel JM, Hoke Jr CH, Sanchez JL, Withers BG, Karabatsos N, et al. Japanese encephalitis vaccine (inactivated, BIKEN) in U.S. soldiers: immunogenicity and safety of vaccine administered in two dosing regimens. American Journal of Tropical Medicine and Hygiene 1999;61:288–93.
- [29] Chunsuttiwat S. Issues related to integration of JE vaccine into national EPI: experience from Thailand. Thailand: Department of Communicable Disease Control, Ministry of Public Health; 1998.
- [30] Chunsuttiwat S. Japanese encephalitis in Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 1989;20:593–7.
- [31] Campbell GL, Hills SL, Fischer M, Jacobson JA, Hoke CH, Hombach JM, et al. Estimated global incidence of Japanese encephalitis: a systematic review. Bulletin of the World Health Organization 2011;89:766–74.
- [32] Innis BL, Nisalak A, Nimmannitya S, Kusalerdchariya S, Chongswasdi V, Suntayakorn S, et al. An enzyme-linked immunosorbent assay to characterize dengue infections where dengue and Japanese encephalitis co-circulate. American Journal of Tropical Medicine and Hygiene 1989;40:418–27.
- [33] Lyons A, Kanesa-thasan N, Kuschner RA, Eckels KH, Putnak R, Sun W, et al. A phase 2 study of a purified, inactivated virus vaccine to prevent Japanese encephalitis. Vaccine 2007;25:3445–53.
- [34] See Section 14: Japanese Encephalitis Vaccine, Inactivated, Adsorbed Package Inserts at http://www.fda.gov/downloads/BiologicsBloodVaccines/Vaccines/ ApprovedProducts/UCM142570.pdf (accessed 18.05.13) and http://www.fda.

- gov/downloads/BiologicsBloodVaccines/Vaccines/ApprovedProducts/UCM123761.pdf (accessed 18.05.13).
- [35] Vaughn DW, Hoke Jr CH. The epidemiology of Japanese encephalitis: prospects for prevention. Epidemiologic Reviews 1992;14: 197–221.
- [36] Endy TP, Nisalak A. Japanese encephalitis virus: ecology and epidemiology. Current Topics in Microbiology and Immunology 2002;267: 11–48.
- [37] Johnson RT, Burke DS, Elwell M, Leake CJ, Nisalak A, Hoke CH, Lorsomrudee W. Japanese encephalitis: immunocytochemical studies of viral antigen and inflammatory cells in fatal cases. Annals of Neurology 1985;18:567–73.
- [38] Burke DS, Ussery MA, Elwell MR, Nisalak A, Leake, Laorakpongse T. Isolation of Japanese encephalitis virus strains from sentinel pigs in northern Thailand, 1982. Transactions of the Royal Society of Tropical Medicine and Hygiene 1985;79:420–1.
- [39] Burke DS, Nisalak A, Lorsomrudee W, Ussery MA, Laorpongse T. Virus-specific antibody-producing cells in blood and cerebrospinal fluid in acute Japanese encephalitis. Journal of Medical Virology 1985;17:283–92.
- [40] Leake CJ, Ussery MA, Nisalak A, Hoke CH, Andre RG, Burke DS. Virus isolations from mosquitoes collected during the 1982 Japanese encephalitis epidemic in northern Thailand. Transactions of the Royal Society of Tropical Medicine and Hygiene 1986;80:831–7.
- [41] Leake CJ, Burke DS, Nisalak A, Hoke CH. Isolation of Japanese encephalitis virus from clinical specimens using a continuous mosquito cell line. American Journal of Tropical Medicine and Hygiene 1986;35:1045–50.
- [42] Libraty DH, Nisalak A, Endy TP, Suntayakorn S, Vaughn DW, Innis BL. Clinical and immunological risk factors for severe disease in Japanese encephalitis. Transactions of the Royal Society of Tropical Medicine and Hygiene 2002;96:173–8.
- [43] Lemon SM, Binn LN. Serum neutralizing antibody response to hepatitis A virus. Journal of Infectious Diseases 1983;148:1033–9.
- [44] Binn LN, Bancroft WH, Lemon SM, Marchwicki RH, LeDuc JW, Trahan CJ, Staley EC, Keenan CM. Preparation of a prototype inactivated hepatitis A virus vaccine from infected cell cultures. Journal of Infectious Diseases 1986;153:749–56.
- [45] Sjogren MH, Hoke CH, Binn LN, Eckels KH, Dubois DR, Lyde L, et al. Immunogenicity of an inactivated hepatitis A vaccine. Annals of Internal Medicine 1991;114:470–1.
- [46] Hoke Jr CH, Binn LN, Egan JE, DeFraites RF, MacArthy PO, Innis BL, et al. Hepatitis A in the US Army: epidemiology and vaccine development. Vaccine 1992;10(Suppl. 1):S75–9.
- [47] Innis BL, Snitbhan R, Hoke CH, Munindhorn W, Laorakpongse T. The declining transmission of hepatitis A in Thailand. Journal of Infectious Diseases 1991:163:989-95.
- [48] Innis BL, Snitbhan R, Kunasol P, Laorakpongse T, Poopatanakool W, Suntayakorn S, et al. Field efficacy trial of inactivated hepatitis A vaccine among children in Thailand. Vaccine 1992;10(S1):S159.
- [49] Innis BL, Snitbhan R, Kunasol P, Laorakpongse T, Poopatanakool W, Kozik CA, et al. Protection against hepatitis A by an inactivated vaccine. Journal of the American Medical Association 1994;271:1328–34.
- [50] See Package Insert, Page 8, Item 14.1 http://us.gsk.com/products/assets/ us.havrix.pdf
- [51] Teppakdee A, Tangwitoon A, Khemasuwan D, Tangdhanakanond K, Suramaethakul N, Sriratanaban J, et al. Cost-benefit analysis of hepatitis A vaccination in Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 2002;33:118–27.
- [52] Participants in the 2001 Conference on Ethical Aspects of Research in Developing Countries. Moral standards for research in developing countries: from "reasonable availability" to "fair benefits". Hastings Center Report 2004:34:17–27.
- [53] Armed Forces Health Surveillance Center (AFHSC). Hospitalizations for hepatitis A, B, and C, active component, U.S. Armed Forces, 1991–2011. MSMR 2012;19(August):18–21.
- [54] Dagan R, Leventhal A, Anis E, Slater P, Ashur Y, Shouval D. Incidence of hepatitis A in Israel following universal immunization of toddlers. Journal of the American Medical Association 2005;294:202–10.
- [55] WHO. Technical guides for diagnosis, treatment, surveillance, prevention and control of Dengue hemorrhagic fever. Geneva: World Health Organization; 1075
- [56] Henchal EA, McCown JM, Seguin MC, Gentry MK, Brandt WE. Rapid identification of dengue virus isolates by using monoclonal antibodies in an indirect immunofluorescence assay. American Journal of Tropical Medicine and Hygiene 1983;32:164–9.
- [57] Henchal EA, Gentry MK, McCown JM, Brandt WE. Dengue virus-specific and flavivirus group determinants identified with monoclonal antibodies by indirect immunofluorescence. American Journal of Tropical Medicine and Hygiene 1982;31:830–6.
- [58] Gentry MK, Henchal EA, McCown JM, Brandt WE, Dalrymple JM. Identification of distinct antigenic determinants on dengue-2 virus using monoclonal antibodies. American Journal of Tropical Medicine and Hygiene 1982;31(3 Pt 1):548–55;
 - Stephens HA, Klaythong R, Sirikong M, Vaughn DW, Green S, Kalayanarooj S, et al. HLA-A and -B allele associations with secondary dengue virus infections correlate with disease severity and the infecting viral serotype in ethnic Thais. Tissue Antigens 2002;60:309–18.

- [59] Wichmann O, Yoon I, Sabchareon A, Vong S, Gibbons RV, Mammen MP, et al. Dengue in Thailand and Cambodia: using data from prospective cohort studies for better disease burden estimates. PLoS Neglected Tropical Diseases 2011:5:e996.
- [60] Kalayanarooj S, Vaughn DW, Nimmannitya S, Green S, Suntayakorn S, Kunentrasai N, et al. Early clinical and laboratory indicators of acute dengue illness. Journal of Infectious Diseases 1997;176:313–21.
- [61] Sudiro TM, Ishiko H, Green S, Vaughn DW, Nisalak A, Kalayanarooj S, et al. Rapid diagnosis of dengue viremia by reverse transcriptase-polymerase chain reaction using 3'-noncoding region universal primers. American Journal of Tropical Medicine and Hygiene 1997;56:424–9.
- [62] Vaughn DW, Green S, Kalayanarooj S, Innis BL, Nimmannitya S, Suntayakorn S, et al. Dengue in the early febrile phase: viremia and antibody responses. Journal of Infectious Diseases 1997;176:322–30.
- [63] Rico-Hesse R, Harrison LM, Nisalak A, Vaughn DW, Kalayanarooj S, Green S, et al. Molecular evolution of dengue type 2 virus in Thailand. American Journal of Tropical Medicine and Hygiene 1998;58:96–101.
- [64] Mathew A, Kurane I, Green S, Stephens HAF, Vaughn DW, Kalayanarooj S, et al. Predominance of HLA restricted cytotoxic T-lymphocyte responses to serotype-cross-reactive epitopes on nonstructural proteins following natural secondary dengue virus infection. Journal of Virology 1998;72:3999–4004.
- [65] Cuzzubbo AJ, Vaughn DW, Nisalak A, Suntayakorn S, Aaskov J, Devine PL. Detection of specific antibodies in saliva during dengue infection. Journal of Clinical Microbiology 1998;36:3737–9.
- [66] Sudiro TM, Ishiko H, Rothman AL, Kershaw DE, Green S, Vaughn DW, et al. Microplate-reverse hybridization method to determine dengue virus serotype. Journal of Virologic Methods 1998;73:229–35.
- [67] Mathew A, Kurane I, Green S, Vaughn DW, Kalyanarooj S, Suntyakorn S, et al. Impaired T cell proliferation in acute dengue infection. Journal of Immunology 1999;162:5607–15.
- [68] Green S, Pichyangkul S, Vaughn DW, Kalayanarooj S, Nimmannitya S, Nisalak A, et al. Early CD69 expression on peripheral blood lymphocytes from children with dengue hemorrhagic fever. Journal of Infectious Diseases 1999;180:1429–35.
- [69] Green S, Vaughn DW, Kalayanarooj S, Nimmannitya S, Suntayakorn S, Nisalak A, et al. Early immune activation in acute dengue illness is related to development of plasma leakage and disease severity. Journal of Infectious Diseases 1999;179:755–62.
- [70] Green S, Vaughn DW, Kalayanarooj S, Nimmannitya S, Suntayakorn S, Nisalak A, et al. Elevated plasma interleukin-10 levels in acute dengue correlate with disease severity. Journal of Medical Virology 1999;59:329–34.
- [71] Vaughn DW, Green S, Kalayanarooj S, Innis BL, Nimmannitya S, Suntayakorn S, et al. Dengue viremia titer, antibody response pattern, and virus serotype correlate with disease severity. Journal of Infectious Diseases 2000;181:2–9.
- [72] Sudiro TM, Zivny J, Ishiko H, Green S, Vaughn DW, Kalayanarooj S, et al. Analysis of plasma viral RNA levels during acute dengue virus infection using quantitative competitor reverse transcription-polymerase chain reaction. Journal of Medical Virology 2001;63:29–34.
- [73] Gagnon SJ, Leporati A, Green S, Kalayanarooj S, Vaughn DW, Stephens HAF, et al. T cell receptor V®□ gene usage in Thai children with dengue virus infection. American Journal of Tropical Medicine and Hygiene 2001;64: 41–8
- [74] Gagnon SJ, Mori M, Kurane I, Green S, Vaughn DW, Kalayanarooj S, et al. Cytokine gene expression and protein production in peripheral blood mononuclear cells of children with acute dengue virus infections. Journal of Medical Virology 2002;67:41–6.
- [75] Zivna I, Green S, Vaughn DW, Kalayanarooj S, Stephens HAF, Chandanayingyong D, et al. The magnitude of T cell response to an HLAB7-restricted epitope on the dengue NS3 protein is associated with the severity of clinical disease in acute dengue virus infection. Journal of Immunology 2002;168:5959–65.
- [76] Stephens HAF, Klaythong R, Sirikong M, Vaughn DW, Green S, Kalayanarooj S, et al. HLA-A and -B allele associations with secondary dengue virus infections correlate with disease severity and the infecting virus serotype in ethnic Thais. Tissue Antigens 2002;60:309–18.
- [77] Vejbaesya S, Luangtrakool P, Luangtrakool K, Kalayanarooj S, Vaughn DW, Endy TP, et al. TNF and LTA gene, allele, and extended HLA haplotype associations with severe dengue virus infection in ethnic Thais. Journal of Infectious Diseases 2009;199:1442–8.
- [78] Srikiatkhachorn A, Gibbons RV, Green S, Libraty DH, Thomas SJ, Endy TP, et al. Dengue hemorrhagic fever: the sensitivity and specificity of the WHO definition in identifying severe dengue cases in Thailand, 1994–2005. Clinical Infectious Diseases 2010;50:1135–43.
- [79] Endy TP, Chunsuttiwat S, Nisalak A, Libraty DH, Green S, Rothman AL, et al. Epidemiology of inapparent and symptomatic acute dengue virus infection: a prospective study of primary school children in Kamphaeng Phet, Thailand. American Journal of Epidemiology 2002;156:40–51.
- [80] Endy TP, Nisalak A, Chunsuttiwat S, Libraty DH, Green S, Rothman AL, et al. Spatial and temporal circulation of dengue virus serotypes: a prospective study of primary school children in KPP, Thailand. American Journal of Epidemiology 2002;156:52–9.
- [81] Mangada MM, Endy TP, Nisalak A, Chunsuttiwat S, Vaughn DW, Libraty DH, et al. Dengue-specific T cell responses in peripheral blood mononuclear cells obtained prior to secondary dengue virus infections in Thai schoolchildren. Journal of Infectious Diseases 2002;185:1697–703.
- [82] Raengsakulrach B, Nisalak A, Maneekarn N, Yenchitsomanus PT, Limsomwong C, Jairungsri A, et al. Comparison of four reverse transcription-polymerase

- chain reaction procedures for the detection of dengue virus in clinical specimens. Journal of Virological Methods 2002;105:219–32.
- [83] Wittke V, Robb TE, Thu HM, Nisalak A, Nimmannitya S, Kalayanrooj S, et al. Extinction and rapid emergence of strains of dengue 3 virus during an interepidemic period. Virology 2002;301:148–56.
- [84] Endy TP, Nisalak A, Chunsuttitwat S, Vaughn DW, Green S, Ennis FA, et al. Relationship of preexisting dengue virus (DV) neutralizing antibody levels to viremia and severity of disease in a prospective cohort study of DV infection in Thailand. Journal of Infectious Diseases 2004;189:990–1000.
- [85] Zhang C, Mammen Jr MP, Chinnawirotpisan P, Klungthong C, Rodpradit P, Monkongdee P, et al. Clade replacements in dengue virus serotypes 1 and 3 are associated with changing serotype prevalence. Journal of Virology 2005;79:15123–30.
- [86] Laoprasopwattana K, Libraty DH, Endy TP, Nisalak A, Chunsuttiwat S, Vaughn DW, et al. Dengue virus (DV) enhancing antibody activity in pre-illness plasma does not predict subsequent disease severity or viremia in secondary DV infection. Journal of Infectious Diseases 2005:192:510–9.
- [87] Clark DV, Mammen Jr MP, Nisalak A, Puthimethee V, Endy TP. Economic impact of dengue fever/dengue hemorrhagic fever in Thailand at the family and population levels. American Journal of Tropical Medicine and Hygiene 2005;72:786–91.
- [88] Myint KS, Endy TP, Mongkolsirichaikul D, Manomuth C, Kalayanarooj S, Vaughn DW, et al. Cellular immune activation in children with acute dengue virus infections is modulated by apoptosis. Journal of Infectious Diseases 2006;194:600-7.
- [89] Koenraadt CJ, Tuiten W, Sithiprasasna R, Kijchalao U, Jones JW, Scott TW. Dengue knowledge and practices and their impact on *Aedes aegypti* populations in KPP, Thailand. American Journal of Tropical Medicine and Hygiene 2006;74:692–700.
- [90] Anderson KB, Chunsuttiwat S, Nisalak A, Mammen MP, Libraty DH, Rothman AL, et al. Burden of symptomatic dengue infection in children at primary school in Thailand: a prospective study. Lancet 2007;369:1452–9.
- [91] Libraty DH, Myint KS, Murray CK, Gibbons RV, Mammen MP, Endy TP, et al. A comparative study of leptospirosis and dengue in Thai children. PLoS Neglected Tropical Diseases 2007;1:111.
- [92] Koenraadt CJ, Jones JW, Sithiprasasna R, Scott TW. Standardizing container classification for immature *Aedes aegypti* surveillance in KPP, Thailand. Journal of Medical Entomology 2007;44:938–44.
- [93] Mammen MP, Pimgate C, Koenraadt CJ, Rothman AL, Aldstadt J, Nisalak A, et al. Spatial and temporal clustering of dengue virus transmission in Thai villages. PLoS Medicine 2008;5:1605–16 [article no. e205].
- [94] Jarman RG, Holmes EC, Rodpradit P, Klungthong C, Gibbons RV, Nisalak A, et al. Microevolution of Dengue viruses circulating among primary school children in KPP, Thailand. Journal of Virology 2008;82:5494–500.
- [95] Lambrechts L, Chevillon C, Albright RG, Thaisomboonsuk B, Richardson JH, Jarman RG, et al. Genetic specificity and potential for local adaptation between dengue viruses and mosquito vectors. BMC Evolutionary Biology 2009;9:160.
- [96] Van Panhuis WG, Gibbons RV, Endy TP, Rothman AL, Nisalak A, Burke DS, et al. Inferring the serotype of dengue virus infections based on pre- and post-infection neutralizing antibody titers. Journal of Infectious Diseases 2010;202:1002–10.
- [97] Endy TP, Yoon IK, Mammen MP. Prospective cohort studies of dengue viral transmission and severity of disease. Current Topics in Microbiology and Immunology 2010;338:1–13.
- [98] Endy TP, Anderson KB, Nisalak A, Yoon I, Green S, Rothman AL, et al. Determinants of inapparent and symptomatic dengue infection in a prospective study of primary school children in KPP, Thailand. PLoS Neglected Tropical Diseases 2011;5:e975.
- [99] Anderson K, Gibbons RV, Thomas SJ, Rothman AL, Nisalak A, Berkelman R, et al. Pre-existing Japanese encephalitis virus (JEV) neutralizing antibodies are associated with an increased risk of symptomatic dengue (DENV) illness in a prospective cohort study of school-children in Thailand. PLoS Neglected Tropical Diseases 2011;5:e1311.
- [100] Aldstadt J, Koenraadt CJ, Fansiri T, Kijchalao U, Richardson J, Jones JW, et al. Ecological modeling of Aedes aegypti (L.) pupal production in rural KPP, Thailand. PLoS Neglected Tropical Diseases 2011;5(1):e940.
- [101] Aldstadt J, Yoon IK, Tannitisupawong D, Jarman RG, Thomas SJ, Gibbons RV, et al. Space-time analysis of hospitalised dengue patients in rural Thailand reveals important temporal intervals in the pattern of dengue virus transmission. Tropical Medicine and International Health 2012;17: 1076–85.
- [102] Yoon IK, Rothman AL, Tannitisupawong D, Srikiatkhachorn A, Jarman RG, Aldstadt J, et al. The hidden threat: mildly symptomatic viremic dengue infections in rural Thai schools and villages. JID 2012;206:389–98.
- [103] Yoon IK, Getis A, Aldstadt J, Rothman AL, Tannitisupawong D, Koenraadt CJM, et al. Fine scale spatiotemporal clustering of dengue virus transmission in children and Aedes aegypti in rural Thai villages. PLoS Neglected Tropical Disease 2012;6:e1730.
- [104] Anderson KB, Endy TP, Mammen MP, Greene S, Yoon IK, Gibbons RV. A shorter time interval between first and second dengue infections is associated with protection from clinical illness in a prospective school-based cohort in Thailand: [under review].
- [105] Rabaa MA, Klungthong C, Yoon IK, Holmes EC, Chinnawirotpisan P, Thaisomboonsuk B, et al. Frequent in-migration and highly focal transmission of dengue viruses among children in KPP, Thailand. PLoS Neglected Tropical Diseases 2012, http://dx.doi.org/10.1371/journal.pntd.0001990.

- [106] Scott TW, Chow E, Strickman D, Kittayapong P, Wirtz RA, Lorenz LH, et al. Blood-feeding patterns of *Aedes aegypti* (Diptera: Culicidae) collected in a rural Thai village. Journal of Medical Entomology 1993;30:922–7.
- [107] Solomon T, Dung NM, Vaughn DW, Kneen R, Thao LT, Raengsakulrach B, et al. Neurological manifestations of dengue infection. Lancet 2000;355: 1053-9
- [108] Jones JW, Sithiprasasna R, Schleich S, Coleman RE. Evaluation of selected traps as tools for conducting surveillance for adult Aedes aegypti in Thailand. Journal of the American Mosquito Control Association 2003;19:148–50.
- [109] Zhang C, Mammen Jr MP, Chinnawirotpisan P, Klungthong C, Rodpradit P, Nisalak A, et al. Structure and age of genetic diversity of dengue virus type 2 in Thailand. Journal of General Virology 2006;87:873–83.
- [110] Myint KS, Gibbons RV, Murray CK, Rungsimanphaiboon K, Supornpun W, Sithiprasasna R, et al. Leptospirosis in KPP, Thailand. American Journal of Tropical Medicine and Hygiene 2007;76:135–8.
- [111] Gibbons RV, Kalanarooj S, Jarman RG, Nisalak A, Vaughn DW, Endy TP, et al. Analysis of repeat hospital admissions for dengue to estimate the frequency of third or fourth dengue infections resulting in admissions and dengue hemorrhagic fever, and serotype sequences. American Journal of Tropical Medicine and Hygiene 2007;77:910–3.
- [112] Thomas SJ, Aldstadt J, Jarman RG, Tannitisupawong D, Yoon IK, Ponlawat A, et al. Dengue virus circulation and virus-vector-host interactions in KPP Province, Thailand; [in preparation].
- [113] Salje H, Lessler J, Endy TP, Curriero F, Gibbons RV, Nisalak A, et al. Revealing the micro-scale spatial signature of dengue transmission and immunity in an urban population. Proceedings of the National Academy of Science 2012:109:9235–8.
- [114] Libraty DH, Myint KS, Murray CK, Gibbons RV, Mammen MP, Endy TP, et al. A comparative study of leptospirosis and dengue in Thai children. PLoS Neglected Tropical Diseases 2007;1(3):e111.
- [115] Pavlin JA, Hickey AC, Ulbrandt N, Chan YP, Endy TP, Boukhvalova MS, et al. Human metapneumovirus reinfection among children in Thailand determined by ELISA using purified soluble fusion protein. Journal of Infectious Diseases 2008;198:836–42.
- [116] Burke RL, Vest KG, Eick AA, et al. Department of defense influenza and other respiratory disease surveillance during the 2009 pandemic. BMC Public Health 2011;11(S2):S6.
- [117] Khuntirat BP, Yoon IK, Blair PJ, Krueger WS, Chittagarnpitch M, Putnam SD, et al. Evidence for subclinical avian influenza virus infections among rural Thai villagers. Clinical Infectious Diseases 2011;53:e107–16.
- [118] Johns MC, Burke RL, Vest KG, et al. A growing global network's role in outbreak response: AFHSC-GEIS 2008–2009. BMC Public Health 2011;11(S2): S3.

- [119] Sanchez JL, Johns MC, Burke RL, et al. Capacity-building efforts by the AFHSC-GEIS program. BMC Public Health 2011;11(S2):S4.
- [120] Sabin AB. Epidemic encephalitis in military personnel; isolation of Japanese B virus on Okinawa in 1945, serologic diagnosis, clinical manifestations, epidemiologic aspects and use of mouse brain vaccine. Journal of the American Medical Association 1947;133:281–93.
- [121] Shrestha MP, Scott RM, Joshi DM, Mammen Jr MP, Thapa GB, Thapa N, et al. Safety and efficacy of a recombinant hepatitis E vaccine. New England Journal of Medicine 2007;356:895–903.
- [122] Myint KS, Endy TP, Shrestha MP, Shrestha SK, Vaughn DW, Innis BL, et al. Hepatitis E antibody kinetics in Nepalese patients. Transactions of the Royal Society of Tropical Medicine and Hygiene 2006;100:938–41.
- [123] Rerks-Ngarm S, Pitisuttithum P, Nitayaphan S, Kaewkungwal J, Chiu J, Paris R, et al. Vaccination with ALVAC and AIDSVAX to prevent HIV-1 infection in Thailand. New England Journal of Medicine 2009;361:2209–20.
- [124] Muangchana C, Henprasertthae N, Nurach K, Theppang K, Yoocharoen P, Varinsathien P, et al. Effectiveness of mouse brain-derived inactivated Japanese encephalitis vaccine in Thai National Immunization Program: a case-control study. Vaccine 2012;30:361-7.
- [125] Siraprapasiri T, Sawaddiwudhipong W, Rojanasuphot S. Cost benefit analysis of Japanese encephalitis vaccination program in Thailand. Southeast Asian Journal of Tropical Medicine and Public Health 1997;28: 143-8
- [126] Communication provided by Dr. Bruce Innis "The last time I was in KPP was 2006. I went looking to purchase some wine for a dinner. The attendant in the store was a young adult, around 25 years of age. We chatted about why I was in KPP and I explained I was escorting colleagues who were considering the feasibility of a dengue vaccine study in KPP. She said she had been in a hepatitis vaccine project 15 years earlier and I explained that I knew something about that and moreover, because of that study, a new vaccine was licensed for use globally. She was pleased, but much more interested to meet one of the foreigners who helped initiate the trial in her youth and to know that perhaps there would be another field trial someday concerning dengue. I was struck by her recall that hepatitis trial made an impression on her (and me too)! I suppose the same is true for participants in all of the major multi-year cohort studies involving children there."
- [127] http://www.ich.org/products/guidelines/efficacy/article/efficacy-guidelines. html (accessed 26.06.13).
- [128] Tuckman BW. Developmental sequence in small groups. Psychological Bulletin 1965;63:384–99.
- [129] Russell PK. Excellence in research is not enough. American Journal of Tropical Medicine and Hygiene 1984;33:319–24.